Ioannis Bantounas

Assistant Professor

Short Curriculum Vitae

Posts held and academic qualifications

2025-Present	**Assistant Professor of Molecular Biology**, Department of Molecular Biology and Genetics, Democritus University of Thrace (DUTh)
2010-2025	**Post-doctoral Research Associate,** Faculty of Biology, Medicine and Health, University of Manchester, UK.
2009-2010	**Post-doctoral Researcher**, Institute of Biological Research and Biotechnology, National Hellenic Research Foundation, Athens, Greece
2006-2009	**Post-doctoral Researcher**, The Henry-Wellcome Laboratories for Integrated Neuroscience and Endocrinology, Faculty of Medicine and Dentistry, University of Bristol, UK.
2000-2004	**PhD**, The Henry-Wellcome Laboratories for Integrated Neuroscience and Endocrinology, Faculty of Medicine and Dentistry, University of Bristol, UK.
1996-1999	**BSc (Honours, 1st Class) in Cell and Molecular Biology**, Department of Biological Sciences, University of Essex, UK.

Research Interests

- Modelling genetic diseases using human pluripotent stem cells
- Regenerative medicine using human pluripotent stem cells
- Control of gene networks by microRNAs
- Viral vectors

Research Funding Awarded

- **5** grants/awards of a total amount of **£273,213**

Prizes – Distinctions

- My paper *Bantounas et al (2018)* was judged as one of the top ten publications of *Stem Cells Reports* for the year 2018 and was a candidate for F1000' (presently '*Faculty Opinions*') on the F1000 platform
- 2 prizes for conference abstracts
- 2 prizes for best performance in the 1st and final year of undergraduate study, University of Essex (UK)

Publications (May 2025)

- **18** publications (**8** as first and/or corresponding author)
- **1096** citations (h-index: **10**) on Google Scholar,
- **746** citations (h-index **10**) on Scopus.
- **46** conference abstracts, of which **4** were published in journals

Teaching

- Participation in **5** undergraduate courses at DUTh
- Co-supervision of **3** PhD theses (2 completed, 1 current).
- Co-supervision of undergraduate and postgraduate projects (2003-present)

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Assistant Professor

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POSTS HELD & RESEARCH

2025-Present

Assistant Professor of Molecular Biology, Department of Molecular Biology and Genetics, Democritus University of Thrace (DUTh)

2023-2025

Post-doctoral Research Associate, Faculty of Biology, Medicine and Health University of Manchester, UK and NIHR Manchester Biomedical Research Centre (Mental Health / Neuroscience) (Supervisors: Prof. Susan J Kimber and Dr Shruti Garg)

Research undertaken:

- *Creating 2D culture and organoid models from human embryonic stem cells (hESCs) and patient induced pluripotent stem cells (iPSCs) for the study of RASopathies in the central nervous system, with an emphasis on Neurofibromatosis Type-1 (NF1)*

Sep-Dec 2022

Post-doctoral Research Associate, Faculty of Biology, Medicine and Health, University of Manchester, UK. (Supervisor: Prof. Martin P Lowe)

Research undertaken:

- *The role of the intermediate filament cytoskeleton (including Vimentin and associated protein GORAB) in the structure and function of the Golgi apparatus*

2014-2022

Post-doctoral Research Associate, Faculty of Biology, Medicine and Health, University of Manchester, UK. (Supervisor: Prof. Susan J Kimber)

Research undertaken:

- *Using human embryonic stem cells (hESCs) and induced pluripotent stem cells (iPSCs) to investigate molecular pathways in kidney development and disease*
- *A novel role for Glutamate Receptors (GluRs) in the kidney*
- *The role of miRNAs in the regulation of molecular pathways during kidney development*
- *Improvement of kidney differentiation protocols by in vivo implantation of differentiating cells*

2010-2013	**Post-doctoral Research Associate,** Faculty of Life Sciences, University of Manchester, UK. (Supervisor: Dr Alan J Whitmarsh) Research undertaken: • *A novel role of JIP1 (a JNK-scaffold protein) in direct regulation of transcription during excitotoxicity.* • *The interplay of JIP1 and AKT in mediating axonal specification and growth*
2009-2010	**Post-doctoral Researcher**, Institute of Biological Research and Biotechnology, National Hellenic Research Foundation, Athens, Greece (Supervisor: Dr Alexander Pintzas) Research undertaken: • *MiRNAs in signalling pathways involved in colon cancer*
2006-2009	**Post-doctoral Researcher**, The Henry-Wellcome Laboratories for Integrated Neuroscience and Endocrinology, Faculty of Medicine and Dentistry, University of Bristol, UK. (Supervisor: Prof. James B Uney) Research undertaken: • *The role of microRNAs in neuronal stress* • *MicroRNAs in neurodegeneration and stem cells* • *Producing Induced Pluripotent Stem Cells by lentiviral transduction of pluripotency factors*
2004-2005	**Microbiology and Biochemistry Laboratory Assistant,** Military National Service, Medical Corps, Greek Army.

ACADEMIC QUALIFICATIONS

2000-2004	**PhD**, The Henry-Wellcome Laboratories for Integrated Neuroscience and Endocrinology, Faculty of Medicine and Dentistry, University of Bristol, UK. Supervisor: Prof. James B Uney Thesis title: *Developing Adenoviral Hammerhead Ribozyme and siRNA Delivery Systems to Study Neuronal Gene Function*
1996-1999	**BSc (Honours, 1st Class) in Cell and Molecular Biology**, Department of Biological Sciences, University of Essex, UK.

FUNDING AWARDED

2021-2022	**Kidneys for Life, Research Grant (£4,900)** Woolf AS, Kimber SJ, Rooney KM, Bantounas I "Treating a genetic kidney disease in a human stem cell model"
2021-2022	**Kidneys for Life, Research Grant (£5,000)** Kimber SJ, Woolf AS, Bantounas I, Rooney KM "Unravelling which kidney cell types are affected in Renal Cyst and Diabetes Syndrome caused by HNF1B mutation using Single Cell RNAseq of stem cell kidney organoids"
2018-2020	**Kidneys for Life, Research Grant (£10,000)** Woolf AS, Kimber SJ, Bantounas I "Studying human kidney malformations in a dish"
2017-2020	**Kidney Research UK, John Feehally – Stoneygate Grant (£251,813)** Kimber SJ, Woolf AS, Bantounas I "Using stem cell technologies to understand human renal tract malformations"
2012	**Faculty of Life Sciences, University of Manchester, Career Development Award (£1,500)**: "Investigating the potential role of the JIP1 scaffold protein in controlling miRNA transcription"

TEACHING

2025-present	**Undergraduate teaching:** • MBG217: Physiology • MBG223: Gene expression • MBG226: Laboratory Course IV: Physiology and Structural Biology • MBG502: Virology • MBG605: Stem Cell and Regenerative Biology
2015-present	**Co-supervision of PhD theses:** 1. Tengku MFS (2019), University of Manchester "Modelling human genetic kidney malformations caused by *HNF1B* mutations using pluripotent stem cells" 2. Rooney KM (2023), University of Manchester. "Investigating Hepatocyte Nuclear Factor 1B associated renal disease in a human kidney organoid model" 3. Almuwallad S. *(current).* "Human pluripotent stem cell-derived kidney organoids (collecting duct) for modelling disease and eventual therapy"
2003-present	**Co-supervision of undergraduate and postgraduate student projects,** on a yearly basis during my work at the Universities of Bristol and Manchester. This included project design, day-to-day experiment design, laboratory supervision and aiding with write-up.

SCIENTIFIC ORGANISATIONS MEMBERSHIP

- **Hellenic Society for Biological Sciences:** Current member (2025-present)
- **British Society for Developmental Biology:** Former member (2022-2024)
- **British Society for Gene and Cell Therapy (BSGCT):** Former member (2015-2018)
- **British Neuroscience Association (BNA):** Former member (2013)

PEER REVIEWING

- **Kidney Research UK:** Grant applications referee (2018-present)
- ***Stem Cell Reports*:** Reviewer
- ***Cell regeneration*:** Reviewer

PRIZES - DISTINCTIONS

2018	The paper *Bantounas et al (2018)* was judged as one of the top ten publications of *Stem Cells Reports* for the year 2018 and was a candidate for F1000' (presently '*Faculty Opinions*') on the F1000 platform
2018	Second prize for conference abstract at the *Renaltract Conference, Manchester, UK*
2014	Prize for best abstract at the *5th Mercia Stem Cell Alliance Meeting, Liverpool, UK*
1999	The Glaxo Prize – for best final undergraduate year performance in class, University of Essex, UK
1997	The Zeneca Prize – for best 1st undergraduate year performance in class, University of Essex, UK

LANGUAGES

Greek	First language.
English	Fluent; Certificate of Proficiency in English, Cambridge University (1995) and Michigan University (1996). BSc and PhD degrees in english, and British citizenship.
German	Fair; Zertifikat Deutsch als Fremdsprache (1994).
French	Fair; classes during secondary education.

PUBLICATIONS

1. **Bantounas I**, Rooney KM, Lopes FM, Tengku F, Woods S, Zeef LAH, Kuba SY, Bates N, Hummelgaard S, Hillman KA, Cereghini S, Woolf AS, Kimber SJ. (2024). Human pluripotent stem-cell derived kidney organoids reveal tubular epithelial pathobiology of heterozygous HNF1B-associated dysplastic kidney malformations. *Stem Cell Rep.* **19(6)**:859-876
2. Vitali T, Sanchez-Alvarez R, Witkos TM, **Bantounas I**, Cutiongco MFA, Dudek M, Yan G, Mironov AA, Swift J, Lowe M. (2023). Vimentin intermediate filaments provide structural stability to the mammalian Golgi apparatus. *J Cell Sci.* **136(20)**:jcs260577
3. Lopes FM, Kimber SJ, **Bantounas I*** (corresponding author) (2021). In situ hybridization of miRNAs in human embryonic kidney and human pluripotent cell-derived kidney organoids. *Bio-Protocol.* **11(17)**:e4150.

4. **Bantounas I*** (corresponding author), Lopes FM, Rooney KM, Woolf AS, Kimber SJ (2021). The miR-199a/214 cluster controls nephrogenesis and vascularisation in a human pluripotent stem cell model. *Stem Cell Rep.* **16(1)**:134-148.

5. Ranjzad P, Jinks J, Salahi AP, **Bantounas I**, Derby B, Kimber SJ, Woolf AS, Wong JKF (2020). Aberrant differentiation of human pluripotent stem cell-derived kidney precursor cells inside mouse vascularized bioreactors. *Nephron.* **144(10)**:509–524.

6. **Bantounas I*** (corresponding author), Silajdžic E, Woolf AS, Kimber SJ. (2020). Formation of mature nephrons by implantation of human pluripotent stem cell-derived progenitors into mice. *Methods Mol Biol.* **2067**:309-322.

7. **Bantounas I**, Ranjzad P, Tengku F, Silajdžić, Forster D, Asselin M, Lewis P, Lennon R, Plagge A, Wang Q, Woolf AS, Kimber SJ (2018). Generation of functioning nephrons by implanting human pluripotent stem cell-derived kidney progenitors. *Stem Cell Rep.* **10(3)**:766-779.

8. Williams DJ, Archer R, Archibald P, **Bantounas I**, *et al.* (2016). Comparability: manufacturing, characterization and controls, report of a UK Regenerative Medicine Platform Pluripotent Stem Cell Platform Workshop, Trinity Hall, Cambridge, 14-15 September 2015. *Regen Med*. **11(5)**:483-92.

9. Keasey MP, Scott HL, **Bantounas I**, Uney JB, Kelly S (2016). MiR-132 is upregulated by ischemic preconditioning of cultured hippocampal neurons and protects them from subsequent OGD Toxicity. *J Mol Neurosci*. **59(3)**:404-10.

10. Dajas-Bailador F*, **Bantounas I* (*Joint First Authors)**, Jones EV, Whitmarsh AJ (2014). Regulation of axon growth by the JIP1-Akt axis. *J. Cell Sci.* **127(Pt 1)**:230-9.

11. Ferraro A, Kontos CK, Boni T, **Bantounas I**, Siakouli D, Kosmidou V, Vlassi M, Spyridakis Y, Tsipras I., Zografos G, Pintzas A (2014). Epigenetic regulation of miR-21 in colorectal cancer: ITGB4 as a novel miR-21 target and a three-gene network (miR-21-ITGB4-PCDC4) as predictor of metastatic tumor potential. *Epigenetics.* **9(1)**:129-41.

12. Joyce T, Oikonomou E, Kosmidou V, Makrodouli E, **Bantounas I**, Avlonitis S, Zografos G, Pintzas A. (2012). A molecular signature for oncogenic BRAF in human colon cancer cells is revealed by microarray analysis. *Curr Cancer Drug Targets.* **12(7)**:873-98.

13. Scott H, Howarth JL, Lee YB, Wong LF, **Bantounas I**, Phylactou LA, Verkade P, Uney JB (2012). Mir-3120 is a mirror microRNA that targets heat shock cognate protein 70 and auxilin and regulates clathrin vesicle uncoating. *J Biol Chem.* **287(18)**:14726-33.

14. Lee Y-B, **Bantounas I**, Lee D-Y, Phylactou L, Caldwell MA, Uney JB. (2009). Twist-1 regulates the miR-199a/214 cluster during development. *Nucleic Acids Res*. **37(1)**:123-8.

15. **Bantounas I**, Uney JB. (2008). The evolution of adenoviral vectors and their application to gene delivery and RNAi in the CNS. In: *Virus Expression Vectors*. Transworld Research Network, ISBN 8178952734

16. Hjiantoniou E, Anayasa M, Nicolaou P, **Bantounas I**, Saito M, Iseki S, Uney JB, Phylactou LA. (2008). Twist induces reversal of myotube formation. *Differentiation*. **76(2)**:182-92.

17. **Bantounas I**, Glover CP, Kelly S, Iseki S, Phylactou LA, Uney JB. (2005). Assessing adenoviral hammerhead ribozyme and small hairpin RNA cassettes in neurons: inhibition of endogenous caspase-3 activity and protection from apoptotic cell death. *J Neurosci Res*. **79(5)**:661-9.

18. **Bantounas I**, Phylactou LA, Uney JB. (2004). RNA interference and the use of small interfering RNA to study gene function in mammalian systems. *J Mol Endocrinol*. **33(3)**:545-57.

- Citations on **Google Scholar: 1096 (h-index, 10)**
  Citations on **Scopus: 746 (h-index, 10)**

  (May 2025)

CONFERENCE CONTRIBUTIONS

1. **44th Annual Conference of the Hellenic Society for Biological Sciences, Ioannina, Greece, (2025)**
   Bantounas I, O'Flaherty J, Cuvertino S, Garg S, Kimber SJ
   Brain organoids and 2D models derived from patient induced pluripotent stem cells for the study of Neurofibromatosis Type-1

12th Mercia Stem Cell Alliance Meeting, Birmingham, UK, (2024)

2. Bantounas I, O'Flaherty J, Cuvertino S, Garg S, Kimber SJ
   Modelling Neurofibromatosis Type-1 in 2D and 3D using human induced pluripotent stem cells

3. Almuwallad SS, Lopes F, Bantounas I, Woolf AS, Kimber SJ
   Kidney-collecting duct organoids derived from human pluripotent stem cells for disease modelling of *HNF1B*-mutation-induced kidney dysfunction.

International Society for Stem Cell Research 2024 Annual Meeting, Hamburg, Germany, (2024)

4. Kimber SJ, Bantounas I, Rooney KM, Lopes FM, Tengku F, Hillman KA, Woolf AS,.
   New insights into disease mechanisms in HNF1B-associated dysplastic kidney malformations using hPSC-derived kidney organoids
5. Almuwallad SS, Lopes F, Bantounas I, Woolf AS, Kimber SJ
   Kidney-collecting duct organoids derived from human pluripotent stem cells for disease modelling of *HNF1B*-mutation-induced kidney dysfunction.

24th European Nephrogenesis Workshop, London, UK, (2024)

6. Bantounas I (speaker) Rooney KM, Lopes FM, Tengku F, Woods S, Zeef LAH, Lin I-H, Kuba SY, Bates N, Hillman KA, Cereghini S, Woolf AS, Kimber SJ.
   Human pluripotent stem cell-derived kidney organoids reveal tubular epithelial pathobiology of heterozygous HNF1B-associated dysplastic kidney malformations
7. Almuwallad SS, Lopes F, Bantounas I, Woolf AS, Kimber SJ
   Kidney-collecting duct organoids derived from human pluripotent stem cells for disease modelling of *HNF1B*-mutation-induced kidney dysfunction.

8. **2024 Global NF Conference, Brussels, Belgium, (2024)**
   Bantounas I, O'Flaherty J, Cuvertino S, Belle M, Garg S, Kimber SJ
   Using hPSC-derived neuronal cultures to study the pathobiology of the brain in RASopathies: The case of Neurofibromatosis Type 1.

9. **43rd Annual Conference of the Hellenic Society for Biological Sciences, Alexandroupoli, Greece, (2024)**
   Bantounas I, Rooney KM, Lopes FM, Tengku F, Woods S, Zeef LAH, Lin I-H, Kuba SY, Bates N, Hillman KA, Cereghini S, Woolf AS, Kimber SJ.
   Human pluripotent stem cell-derived kidney organoids reveal tubular epithelial pathobiology of heterozygous *HNF1B*-associated dysplastic kidney malformations

11th Mercia Stem Cell Alliance Meeting, Nottingham, UK, (2023)

10. Bantounas I (Speaker), Rooney KM, Lopes FM, Tengku F, Woods S, Zeef LAH, Lin I-H, Bates N, Hillman KA, Cereghini S, Woolf AS, Kimber SJ. Probing disease mechanisms in HNF1B-associated dysplastic kidney malformations with hPSC-derived kidney organoids.
11. Almuwallad S, Lopes FM, Bantounas I, Woolf AS, Kimber SJ.
    Kidney-collecting duct organoids derived from human pluripotent Stem cells for disease modelling and eventual therapy (HNF1B-Mutant Collecting Duct Kidney Organoids)
12. Lopes FM, Nash A, Almuwallad S, Bates N, Bantounas I, Woolf AS, Kimber SJ.
    Advanced human pluripotent stem cell kidney organoid model for investigating development and disease.

13. **Manchester Regenerative Medicine Network and Manchester Rare Conditions Centre Symposium, Manchester, UK, (2023)**
    Bantounas I (Speaker), Rooney KM, Lopes FM, Tengku F, Woods S, Zeef LAH, Kuba SY, Bates N, Hummelgaard S, Hillman KA, Cereghini S, Woolf AS, Kimber SJ.
    Human pluripotent stem-cell derived kidney organoids reveal tubular epithelial pathobiology of heterozygous HNF1B-associated dysplastic kidney malformations.

14. **10th Mercia Stem Cell Alliance Meeting, York, UK, (2022)**
Rooney KM, Bantounas I, Lopes FM, Zeef L, Woolf AS, Kimber SJ.
Investigating HNF1B renal disease in a human kidney organoid model.

UK Kidney Week 2022, Birmingham, UK, (2022)

15. Bantounas I, Tengku F, Rooney KM, Bates N, Woods S, Hillman KA, Woolf AS, Kimber SJ. Using personalised pluripotent stem cell technology to study inherited genetically-defined renal dysplasia.
Bantounas I, Rooney KM, Lopes FM, Zeef LAH, Woolf AS, Kimber SJ.
16. Searching for druggable targets in HNF1B mutant kidney organoids.

British Society for Cell Biology/British Society for Developmental Biology Joint Annual Spring Meeting, University of Warwick, UK, (2022)

17. Bantounas I, Rooney KM, Tengku F, Bates N, Lopes FM, Woods S, Kuba SY, Hillman KA, Woolf AS, Kimber SJ.
IPSC and hESC-derived kidney organoid models to study human kidney development and disease.
18. Rooney KM, Bantounas I, Lopes FM, Zeef LAH, Woolf AS, Kimber SJ.
Investigating *HNF1B* renal disease in a human kidney organoid model.

19. **North of England Developmental Biology Symposium 2021, (2021)**
Bantounas I (speaker), Lopes FM, Rooney KM, Woolf AS, Kimber SJ. The miR-199a/214 cluster controls nephrogenesis and vascularisation in a human pluripotent stem cell model

20. **9th Mercia Stem Cell Alliance Meeting, Chester, UK, (2019)**
Bantounas I, Lopes FM, Woolf AS, Kimber SJ.
The miR-199a/214 cluster controls nephron morphogenesis in a human pluripotent stem cell model of kidney development

21. **22nd European Nephrogenesis Workshop, London, UK, (2019)**
Bantounas I (speaker), Lopes FM, Tengku F, Woolf AS, Kimber SJ.
Using human pluripotent stem cell models to decipher the role of miRNA-199a/214 in epithelial/mesenchymal balance and glomerular differentiation during kidney development

8th Mercia Stem Cell Alliance Meeting, Birmingham, UK, (2018)

22. Bantounas I, Tengku F, Woolf AS, Kimber SJ.
The miR-199a/214 cluster controls the epithelial/mesenchymal balance in a human pluripotent stem cell model of kidney differentiation
23. Tengku F, Bantounas I, Hillman KA, Ranjzad P, Woods S, Bates N, Woolf AS, Kimber SJ.
Modelling genetic kidney malformations using human induced pluripotent stem cell-derived organoids

Renaltract Conference (and 21st Nephrogenesis Workshop), Manchester, UK, (2018)

24. Bantounas I, Tengku F, Woolf AS, Kimber SJ.
Investigating the role of the miRNA-199a/214 cluster in kidney development using a human pluripotent stem cell model
25. Ranjzad P, Bantounas I, Salahi AP, Kimber SJ, Woolf AS, Wong J.
Towards enhancing the vasculature of mini-kidneys formed from human pluripotent stem cells

26. Tengku F, Bantounas I, Woods S, Hillman KA, Ranjzad P, Woolf AS, Kimber SJ.
Modelling genetic kidney malformations using human induced pluripotent stem cell-derived kidney organoids

27. **20th UK/EU Nephrogenesis Workshop, The Roslin Institute, University of Edinburgh, (2017)**
Bantounas I (Speaker). In vivo generation of functional kidney structures using pluripotent stem cell-derived progenitors.

28. **BSGCT Annual Conference / UK Regenerative Medicine Platform Joint Meeting, Cardiff, UK, (2017)**
Bantounas I (speaker), Ranjzad P, Tengku F, Silajdžić E, Lewis P, Wang Q, Plagge A, Forster D, Asselin M, Woolf AS, Kimber SJ.
Regenerating the kidney using lentivirus-labelled human pluripotent stem cells.

29. **7th Mercia Stem Cell Alliance Meeting, Manchester, UK, (2016)**
Bantounas I, Tengku MFS, Silajdzic E, Lewis P, Wang Q, Ranjzad P, Plagge A, Forster D, Asselin M-C,Willams S, Woolf AS, Kimber SJ.
Lentivirus-mediated engineering of human pluripotent stem cells and their kidney derivatives to track cell fates *in vivo*.

30. **The Inaugural UK Regenerative Medicine Conference, London, UK, (2016)**
Bantounas I, Tengku MFS, Silajdzic E, Lewis P, Wang Q, Asselin MC, Woolf AS, Kimber SJ.
Lentivirus-mediated engineering of human pluripotent stem cells and their kidney derivatives to track cell fates *in vivo*.

31. **6th Mercia Stem Cell Alliance Meeting, Sheffield, UK, (2015)**
Bantounas I, Ranjzad P, Linnik I, Woolf AS, Kimber SJ.
Tracing human embryonic stem cell differentiation into kidney cell types using lentiviral and CRISPR-mediated labelling.

32. **Nephrotools 3rd International Conference, Liverpool, UK, (2015)**
Bantounas I, (speaker), Ranjzad P, Woolf AS, Kimber SJ.
Lentivirus- and CRISPR/Cas9-mediated labelling of pluripotent stem cells to trace differentiation into kidney cell types and their localisation *in vivo*.

33. **British Society for Gene and Cell Therapy Annual Meeting, Glasgow, UK, (2015)**
Bantounas I, Ranjzad P, Woolf AS, Kimber SJ.
Using lentiviral vector and CRISPR/Cas9 technology to trace human embryonic stem cell differentiation into kidney cell types.

34. **UK Regenerative Medicine Platform Scientific Showcase, London, UK, (2015)**
Bantounas I (speaker)
Tracing differentiation of human embryonic stem cells into kidney cell types.

35. **5th Mercia Stem Cell Alliance Meeting, Liverpool, UK, (2014)**
Bantounas I, Kimber SJ.
CRISPR/Cas9-mediated reporter gene knock-in in human embryonic stem cells to mark steps along the kidney differentiation pathway.

36. **Nephrogenesis Workshop, Manchester, UK, (2014)**
Bantounas I (speaker)
CRISPR/Cas9-mediated genomic modification of hESC to mark steps along the kidney differentiation pathway.

37. **BNA 2013: Festival of Neuroscience (British Neuroscience Association), London, UK, (2013)**
Bantounas I, Dajas-Bailador F, Whitmarsh AJ.
Neuronal activity regulates axon growth via the JIP1-AKT axis.

38. **21st Meeting of the European Association for Cancer Research, Oslo, Norway, (2010)**
Pintzas A, Joyce T, Oikonomou E, Makrodouli E, Medico E, Cantarella D, Bantounas I.
Bioinformatic analysis of BRAFV600E vs RASG12V signatures in colon cancer cells reveals differential regulation of cellular pathways related to MSI or EMT. Published in: *European Journal of Cancer Supplements* **8(5):** 203.

39th Annual Meeting of the Society for Neuroscience, Chicago, IL, USA, (2009)

39. Keasey M, Bantounas I, Uney JB, Kelly S.
MiR-132 Protects Neurons from Ischaemic Insult.

40. Strickland I, Richards L, Holmes FE, Bantounas I, Uney JB, Wong L-F
Regulation of neurite outgrowth by microRNAs in dorsal root ganglion neurons.

41. **38th Annual Meeting of the Society for Neuroscience, Washington D.C., USA, (2008)**
Bantounas I, Lee Y-B, Howarth JL, Verkade P, Uney JB.
MiR-214 regulates Hsc70 expression and clathrin uncoating in response to heat stress in cortical neurons.

42. **2nd Annual Meeting of the Bristol Neuroregeneration Group, University of Bristol, UK, (2008)**
Bantounas I, Invited Speaker;
MicroRNAs and cell stress: MiR-214 inhibition of Hsc70 via TIAF1.

43. **1st Annual Meeting of the Bristol Neuroregeneration Group, University of Bristol, UK, (2007)**
Bantounas I, Invited Speaker
Investigating the role of microRNAs as regulators of neuronal gene function.
→ Bantounas I, Session chair: Session Title: *In vitro* methods for the study of neurological disease and regeneration.

44. **6th International Congress of Neuroendocrinology, Pittsburgh, PA, USA, (2006)**
Bantounas I, Invited Speaker;
Practical approaches to gene silencing.

45. **British Neuroscience Association, 17th Annual Meeting, Harrogate, UK, (2003)**
Bantounas I, Glover CPJ, Phyalctou LA, Uney JB.
Adenoviral Delivery Of Hammerhead Ribozymes As A Tool To Study Neuronal Gene Function.

46. **5th Annual Meeting of the American Society for Gene Therapy, Boston, MA, USA, (2002)**
Bantounas I, Glover CPJ, Heywood DJ, Hobson RJ, Phylactou LA, Uney JB.
Assessing the Efficacy of Using Hammerhead Ribozymes to Study Neuronal Gene Function. Published in: Molecular Therapy **5(5)** (supplement): S137.